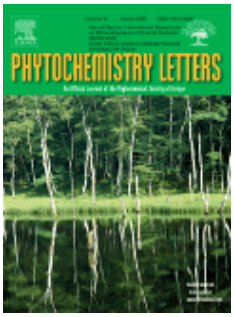
PHYTOCHEMISTRY LETTERS

AUTHOR INFORMATION PACK

TABLE OF CONTENTS

- **Description** **p.1**
- **Impact Factor** **p.1**
- **Abstracting and Indexing** **p.1**
- **Editorial Board** **p.2**
- **Guide for Authors** **p.4**



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Phytochemistry Letters invites rapid communications on all aspects of natural product research including:

- Structural elucidation of natural products
- Analytical evaluation of herbal medicines
- Clinical efficacy, safety and pharmacovigilance of herbal medicines
- Natural product biosynthesis
- Natural product synthesis and chemical modification
- Natural product metabolism
- Chemical ecology
- Biotechnology
- Bioassay-guided isolation
- Pharmacognosy
- Pharmacology of natural products
- Metabolomics
- Ethnobotany and traditional usage
- Genetics of natural products

Manuscripts that detail the isolation of just one new compound are not substantial enough to be sent out of review and are out of scope. Furthermore, where pharmacology has been performed on one new compound to increase the amount of novel data, the pharmacology must be substantial and/or related to the medicinal use of the producing organism.

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Submissions from any field of natural product research are encouraged and the following list is not exhaustive:
• Structural elucidation of natural products
• Analytical evaluation of herbal medicines
• Clinical efficacy, safety and pharmacovigilance of herbal medicines
• Natural product biosynthesis
• Natural product synthesis and semi-synthesis
• Chemical ecology
• Biotechnology
• Bioassay-guided isolation
• Pharmacognosy
• Pharmacology of natural products
• Metabolomics

• Ethnobotany and traditional usage
• Natural product metabolism
• Genetics of natural products

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In the context of Phytochemistry Letters taxonomic issues need to be addressed in a variety of ways.

•It is general practice that voucher specimens should be deposited in a recognized herbarium. These voucher specimens need to be fully cited within the article (with collector, collector number and herbarium). In the case of lesser known plants, we encourage authors to include electronic scans of the specimens as part of their supplementary data.
•As an essential step, authors will have to check the taxonomic validity of the plant names using one of the international databases, and preferably www.theplantlist.org
•In future, such a check will be built into the submission and review process and authors will only be able to submit manuscripts, after the validation of the species' taxonomy.
•A particular problem are complex preparations, especially those containing plant extracts. Here detailed evidence on the authentication during the production needs to be ascertained. In addition fingerprints of the preparations tested are advisable.
•Very commonly these questions have been ignored in clinical studies of herbal preparations. The following two papers make it clear that a correct taxonomic nomenclature is an essential requirement in such studies:
Heinrich and Verpoorte, J. Ethnopharmacol. 2014, doi.org/10.1016/j.jep.2014.01.016 and Rivera et al J. Ethnopharmacol. 2014 http://dx.doi.org/10.1016/j.jep.2013.12.022

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State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

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J. Mabberley, 2nd ed., June 1997, Cambridge University Press; ISBN:0521414210), and preferably be in the form recommended by the International Code of Botanical Nomenclature (http://www.bgbm.fu-berlin.de/iapt/nomenclature/code/tokyo-e/default.htm). Named varieties of cultivars are given e.g. *Lactuca sativa cv.* Grand Rapids. (The official printed version of the International Code of Botanical Nomenclature has been published as International Code of Botanical Nomenclature {Tokyo Code}. Regnum Vegetabile 131. Koeltz Scientific Books, Konigstein. ISBN: 3-87429-367-X or 1-878762-66-4 or 80-901699-l-0.)

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Subsections on the Experimental Procedures should be italicized and inserted as part of the first line of the text to which they apply. Phytochemistry Letters encourages an extensive use of abbreviations (these are below, or the reader is referred to other sources). The Experimental should begin with a subsection entitled General Experimental Procedures. This subsection will typically contain brief details of instruments used, and identification of sources of specialized chemicals, biochemicals and molecular biology kits.

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The Experimental Procedures employed should be concise but sufficiently detailed that a qualified researcher will be able to repeat the studies undertaken, and these should emphasize either truly new procedures or essential modifications of existing procedures. Experimental details normally omitted include: (1) method of preparation of common chemical and biochemical derivatives, (2) excessive details of separation of compounds, proteins and enzymes, e.g. preparation of columns, TLC plates, column and fraction size.

Compound characterization: Physical and spectroscopic data for **new** compounds must be comprehensive, and follow the order shown below: compound name (and assigned number in text); physical state of compound (e.g. oil, crystal, liquid, etc.), melting and/or boiling point; optical rotation and/or circular dichroism measurements, if optically active; UV; IR; 1H NMR; 13C NMR; MS. For all new compounds, either high-resolution mass spectral or elemental analysis data are required. See later section for method of data presentation.

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Elemental analysis results for compounds which have been adequately described in the literature must be given in the form: (Found: C, 62.9; H, 5.4. Calc. for C13H13O4N: C, 63.2; H, 5.3%.) New compounds must be indicated by giving analytical results in the form: (Found: C, 62.9; H, 5.4. C13H13O4N requires: C, 63.2; H, 5.3%.) Thin-layer chromatography
(a) For analytical TLC, dimensions of the plates can be deleted if layer thickness is 0.25 mm.
(b) Abbreviate common adsorbents: (but use silica gel, not SiO2 as this does not describe the material accurately), Al2O3 (alumina).
(c) Preparative forms of the technique should include details of (i) layer thickness (preparative TLC only), (ii) amount of sample applied to the layer, (iii) method of detection used to locate the bands and (iv) the solvent used to recover the compounds from the adsorbent after development.
(d) Special forms of TLC on impregnated adsorbents can be abbreviated, e.g. AgNO3-silica gel (1:9), by wt can be assumed.

Gas chromatography
(a) Detector used should be specified, e.g. dual FID, EC, etc.
(b) Carrier gas and flow rate should be given, e.g. N2 at 30 ml min-1.
(c) Operating conditions, such as injector and detector heater temperatures etc., should be included.
(d) Packed columns, e.g. 6 m x 3 mm (i.d. measurement only) packed with 1% SE-30 (support material and mesh size can be omitted unless unusual).
(e) Capillary columns should be specified, e.g. WCOT (wall coated open tubular), SCOT (support coated open tubular). The split ratio used in the injection system and the injection volume for the sample should also be included.

High performance liquid chromatography
(a) Solvent or solvent gradients used together with flow rate should be given.
(b) Column dimensions (length x i.d. only) and packing used.
(c) Method of detection employed, e.g. UV or refractive index.

Biochemical conventions

Unless a common biochemical term (e.g. ATP, NADH), biochemicals that are abbreviated should be spelled out in full (in brackets) immediately following their first usage in the text. Enzyme names are typically not abbreviated, unless there are accepted abbreviations, such as ATPase. Where possible, E.C. numbers should be used for enzymes, and the recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology (IUBMB) should be used (see below).
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(b) pH optima should be given together with pH values for half maximal activity.
(c) Kinetic parameters should be expressed as Vmax, Km etc.
(d) Enzyme inhibitors-effectiveness should be expressed as Ki or concentration for half-maximal activity.
(e) Optimal temperature of enzymes should not be given. This should be expressed in terms of "Energy of Activation" and "Energy of Activation for Denaturation".
(f) Enzyme nomenclature is now given in "Enzyme Nomenclature, Recommendations", Academic Press (1992) (http://www.chem.qmul.ac.uk/iubmb).

(g) Labelling of proteins and nucleic acids-use of labelled precursors in assessing the rate of synthesis of macromolecules must be validated by evidence of real, direct incorporation. The possibility of occlusion or adsorption of isotopic material should be noted and it should be shown that the labelled precursor is incorporated without prior catabolism.

Protein and nucleotide sequences

The Experimental must contain explicit documentation of the ends of nucleotide probes used in the study if previously unpublished, or by appropriate reference to published nucleotide numbers and/or restriction map.

In manuscripts to be published in Phytochemistry Letters, any new protein and/or nucleotide sequence must have been submitted to EMBL, GenBank or DNA Data Bank of Japan databases, with designated accession number(s) obtained **prior** to paper acceptance by the Regional Editor. The Author(s) must ensure access to this database information by timely release of data prior to publication, as well as providing necessary documentation to those already in the databases.

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About, approximately: ca.
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Dry weight: dry wt; fresh weight: fr. wt
Electricity: V, mA, eV
Force due to gravity (centrifugation): g; rpm (revolutions min-1)
Gas chromatography: GC
Gas chromatography mass spectrometry: GC MS trimethylsilyl derivative: TMSi (TMS cannot be used as this refers to the internal standard tetramethylsilane used in 1H NMR)
High performance liquid chromatography: HPLC
Infrared spectrophotometry: IR
Length: nm, m, mm, cm, m
Literature: lit.
Mass spectrometry: m/z [M]+ (molecular ion, parent ion)
Melting points: uncorr. (uncorrected)
Molecular mass: Da (daltons), kDa
Molecular weight: Mr
Nuclear magnetic resonance: 1H NMR, 13C NMR, Hz, ð
Numbers: e.g. 1, 10, 100, 1000, 10,000: per or -1
Optical rotatory dispersion: ORD
Paper chromatography: PC
Precipitate: ppt.
Preparative thin-layer chromatography: prep. TLC
Radioactivity: dpm (disintegrations per min), Ci (curie), sp. act (specific activity), Bq (1 becquerel=1 nuclear transformation sec-1)
Repetitive manipulations: once, twice, x3, x4, etc.

RRt (relative retention time), Rt (Kovat's retention index), ECL (equivalent chain length term frequently used in fatty acid work)
Saturated: satd.
Solution: soln.
Solvent mixtures including chromatographic solvents: abbreviate as follows n-BuOH HOAc H2O (4:1:5)
Statistics: LSD (least significant difference), s.d. (standard deviation), s.e. (standard error)
Temperature: (with centigrade), mp, mps, mmp, bp
Temperature: temp.
Thin-layer chromatography: TLC, Rf
Time: s, min, h, day, week, month, year
Ultraviolet spectrophotometry: UV, A (absorbance, not OD optical density)
Volume: l (litre), l, ml
Weight: wt, pg, ng, g, mg, g, kg

Inorganics, e.g. AlCl3 (aluminum chloride), BF3 (boron trifluoride), Cr-, CO2, H2, HCl, HClO4 (perchloric acid), HNO3, H2O, H2O2, H2SO4, H3BO3 (boric acid), He, KHCO3 (potassium bicarbonate), KMnO4 (potassium permanganate), KOH, K-Pi buffer (potassium phosphate buffer), LiAlH4 (lithium aluminium hydride), Mg2+, MgCl2, N2, NH3, (NH4)2SO4, Na+, NaBH4 (sodium borohydride), NaCl, NaIO4 (sodium periodate), NaOH, Na2SO3 (sodium sulphite), Na2SO4 (sodium sulphate), Na2S2O3 (sodium thiosulphate), O2, PPi (inorganic phosphate), SO, Tris (buffer).

Organics, e.g. Ac2O (acetic anhydride), n-BuOH (butanol), C6H6 (benzene), CCl4 (carbon tetrachloride), CH2Cl2 (methylene chloride), CHCl3 (chloroform), CH2N2 (diazo-methane), CM (carboxymethyl), DEAE (diethylaminoethyl), DMF (dimethylformamide), DMSO (dimethyl sulphoxide), EDTA (ethylene-diaminetetra-acetic acid), Et2O (diethyl ether), EtOAc (ethyl acetate), EtOH (ethanol), HCO2H (formic acid), HOAc (acetic acid), iso-PrOH (iso-propanol), Me2CO (acetone), MeCOEt (methyl ethyl ketone), MeOH (methanol), NaOAc (sodium acetate), NaOMe (sodium methoxide), petrol (not light-petroleum or petroleum ether), PhOH (phenol), PrOH (propanol), PVP (polyvinylpyrrolidone), TCA (trichloroacetic acid), TFA (trifluoroacetic acid), THF (tetrahydrofuran). 1H NMR solvents and standards: CDCl3 (deuterochloroform), D2O, DMSO-d6 [deuterodimethylsulphoxide, not (CD3)2SO], pyridine-d5 (deuteropyridine), TMS (tetramethylsilane).

For further terms used in biochemistry and molecular biology the authors should see the websites of the nomenclature committees (http://www.chem.qmul.ac.uk/iubmb/).

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